

Kynurenine metabolites of tryptophan: Implications for neurologic diseases

Andrew Freese, BA; Kenton J. Swartz, BA; Matthew J. During, MD, FRACP; and Joseph B. Martin, MD, PhD

Article abstract—Over the past 2 decades, a number of studies have demonstrated that amino acids act as precursors for the biosynthesis of a variety of neuroactive compounds, including catecholamines and indoleamines. For example, the aromatic amino acid L-tryptophan is a precursor for serotonin biosynthesis. Based on this observed precursor relationship, dietary tryptophan supplementation is used to treat a number of neurologic disorders attributed to alterations in serotonergic neurotransmission. Recent studies have revealed that, in addition to serotonin, a number of neuroactive compounds, the kynurenines, are metabolites of tryptophan. Of these, perhaps the most important is quinolinic acid, a neurotoxin that acts at the N-methyl-D-aspartate (NMDA) receptor and whose precursor responsiveness to tryptophan far exceeds that of serotonin. In the central nervous system, kynurenines, and in particular quinolinic acid, may modulate excitatory amino acid transmission, and may act as neurotoxic agents implicated in the pathogenesis of several neurologic diseases.

NEUROLOGY 1990;40:691-695

Tryptophan and brain serotonin. The precursor relationship of plasma and brain tryptophan to serotonin^{1,2} has led to therapeutic interventions based on oral tryptophan supplementation in several neuropsychiatric and related disorders. Included among these are depression, insomnia, aggression, obesity, and hypertension.³⁻⁹ As a result of these speculations, self-administration of tryptophan bought at health food and other stores is now common.

Tryptophan and brain kynurenines. Although tryptophan is the precursor for serotonin biosynthesis, it is now known that its metabolism in brain is more complex. A number of studies have demonstrated that increased plasma (and therefore, brain) levels of tryptophan not only augment brain levels of serotonin, but also the biosynthesis of the kynurenines.¹⁰⁻¹⁶ Of the kynurenines, quinolinic acid may be the most relevant to neurodegenerative diseases, since this dicarboxylic acid has recently been implicated in the etiology of a number of neurologic disorders, including Huntington's disease, temporal lobe epilepsy, glutaric aciduria, and hepatic encephalopathy and coma.¹⁷⁻²³ Studies have demonstrated^{12,13} that tryptophan loading in rats elevates brain tissue levels of quinolinic acid; concentrations of quinolinic acid in rat brain were elevated several-fold by a single injection of 300 mg/kg tryptophan. We have recently shown¹⁵ that a single intraperitoneal injection of tryptophan (250 mg/kg) could elevate *extracellular fluid* levels of quinolinic acid in the brain approximately 200-fold. More physiologic doses of tryptophan (12.5 mg/kg, 50 mg/kg, and 100 mg/kg) led to increases of quinolinic acid of approximately 3-

fold, 40-fold, and 80-fold, respectively, in the brain extracellular fluid compartment.¹⁶ These same doses of tryptophan increased brain extracellular fluid levels of serotonin and its metabolites to only a minor extent (less than 2-fold at a dose of 100 mg/kg tryptophan). The levels of quinolinic acid obtained in these studies at the higher doses of tryptophan are equivalent to those shown to be toxic to neurons *in vitro*.²⁴

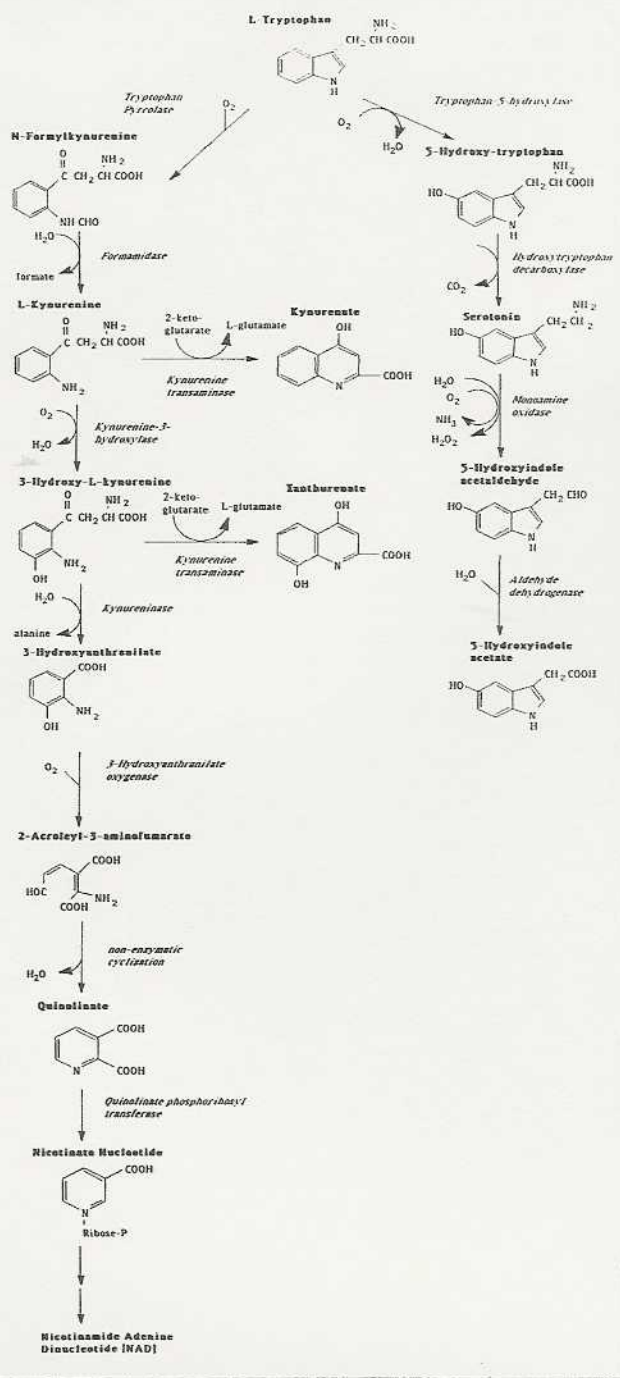
Quinolinic acid and the NMDA (AA₁) receptor. Quinolinic acid toxicity in the brain appears to be mediated through its action as an agonist of the N-methyl-D-aspartate (NMDA) excitatory amino acid receptor.^{25,26} Excitatory amino acid receptor subtypes in mammalian brain have been classified according to their respective affinities for structural analogues of glutamate: N-methyl-D-aspartate (AA₁), quisqualate (AA₂), and kainate (AA₃).^{27,28} Unlike these analogues, quinolinic acid is an endogenous compound in human tissues, making its potential toxic action in the nervous system particularly relevant. Neurotoxicity induced by quinolinic acid occurs preferentially in the neocortex, striatum, and hippocampus, sites in the brain particularly affected by the CNS disorders mentioned above.^{26,29} After injection directly into the brain, quinolinic acid produces axon-sparing lesions³⁰ similar to the pathology found in these disorders. Several antagonists to the NMDA receptor are presently under review as therapies for these and other neurologic disorders.³¹⁻³⁴

Regulation of tryptophan and quinolinate metabolism. As shown in the figure, quinolinic acid is formed from a precursor, 3-hydroxyanthranilate, by the

From the Department of Neurology (A. Freese and K.J. Swartz, and Drs. During and Martin), Massachusetts General Hospital and Harvard Medical School, Boston, MA; the Division of Health Sciences and Technology (A. Freese), Massachusetts Institute of Technology, Cambridge, MA; Program in Neuroscience (K.J. Swartz), Harvard Medical School, Boston, MA; and the Departments of Pharmacology and Psychiatry (Dr. During), Yale University School of Medicine, New Haven, CT. Dr. Martin's current address is Office of the Dean, University of California, San Francisco, CA.

Received April 28, 1989. Accepted for publication in final form September 13, 1989.

Address correspondence and reprint requests to Andrew Freese, Neurology Research, Edwards 4, Massachusetts General Hospital, Fruit Street, Boston, MA 02114.



acid, QPRT, might be unable to remove. This possibility is supported by our findings that increases in quinolinic acid biosynthesis after a tryptophan load lead to markedly elevated extracellular fluid levels of the neurotoxin.^{35,16}

Plasma, and hence brain, levels of tryptophan are regulated precisely. Human serum albumin binds tryptophan,^{38,39} and the availability of tryptophan to cross the blood-brain barrier is determined by the amount of plasma free tryptophan. The albumin binding is saturable; therefore, elevated tryptophan intake can lead to elevated plasma levels of free tryptophan.⁴⁰ Furthermore, access of tryptophan across the blood-brain barrier is controlled by a specific large neutral amino acid transport system that transports all large neutral amino acids, including tryptophan, tyrosine, phenylalanine, valine, and isoleucine.⁴¹ Thus, access of tryptophan to this carrier is determined by the ratio of levels of free tryptophan to all other neutral amino acids in plasma, rather than the level of free tryptophan in plasma alone.^{38,42}

Tryptophan and quinolinic acid in neurologic disorders. Huntington's disease. In Huntington's disease, an autosomal dominant genetic disorder marked by choreoathetosis, a number of studies have suggested that a disorder in tryptophan metabolism may be present. Plasma levels of free tryptophan (versus bound to albumin) were elevated in 1 study⁴³; in others, levels of other large neutral amino acids were decreased in the fasting state in Huntington's disease compared with controls.^{44,45} Belendiuk's group⁴³ also demonstrated a direct correlation between plasma free tryptophan levels and the severity of chorea. Another study⁴⁶ found that daily feeding of tryptophan to juvenile Huntington's patients worsened their chorea, although the results of this study remain controversial.⁴⁷⁻⁴⁹ Of considerable importance is the recent finding⁵⁰ that the brains of Huntington's disease patients have a 3- to 4-fold increase in the activity of the quinolinic acid-synthesizing enzyme, 3HAO, in the corpus striatum, the area in the brain most affected by this disease process. In contrast, the activity of the quinolinic acid-degrading enzyme, QPRT, is relatively unaffected in Huntington's disease.^{37,51} All these studies suggest that Huntington's disease may prove to be caused by a disorder of the metabolism of tryptophan via the kynurenine pathway.

In addition, we have shown that when quinolinic acid is injected into the corpus striatum of rats and primates, a pattern of cell death results that is similar to that found in postmortem striatum of Huntington's disease victims.²³ In both the quinolinic acid-lesioned model and Huntington's disease, striatal levels of substance P-like immunoreactivity and gamma-aminobutyric acid are reduced, whereas levels of somatostatin-like immunoreactivity and neuropeptide Y-like immunoreactivity are spared, and in some cases increased.^{23,52,53}

Another disorder that results in altered tryptophan metabolism, glutaric aciduria, also causes pathologic de-

Figure. The tryptophan metabolic pathway. The metabolism of tryptophan splits between the indoleamine pathway producing serotonin and the kynurenine pathway producing quinolinic and kynurenic acids.

enzyme 3-hydroxyanthranilate oxygenase (3HAO); in turn, quinolinic acid is catabolized by the enzyme quinolinic acid phosphoribosyl transferase (QPRT).³⁵ Of particular importance is the finding that the maximal enzymatic rate (Vmax) for 3HAO is approximately 80-fold higher than that for QPRT, although the Michaelis-Menten constant (Km) for both enzymes is about the same.^{36,37} Thus, an increased flux of tryptophan into the kynurenine pathway could increase levels of quinolinic

struction of the corpus striatum and the clinical manifestation of choreoathetosis; in an animal model of glutaric aciduria, brain quinolinic acid levels were elevated.¹²

Hepatic disorders. Many severe hepatic disorders are associated with neurologic sequelae. Such effects constitute "hepatic encephalopathy," which can result from alcoholic cirrhosis, chronic hepatitis, or other liver disorders. Although the biochemical alterations responsible for hepatic encephalopathy remain poorly defined, 2 of the most consistent findings in both clinical cases and animal models are elevated blood and brain levels of ammonia and tryptophan.⁵⁴⁻⁶⁰ Interestingly, ammonia facilitates the transport of tryptophan across the blood-brain barrier.⁶¹ Liver failure is also associated with a marked increase in the plasma concentrations of unesterified free fatty acids, which compete with tryptophan for albumin binding sites.⁶² Thus, increases in unesterified free fatty acids can increase plasma levels of free tryptophan, and as a result its access to the brain. In addition, hypoalbuminemia that can result from liver disorders would liberate otherwise bound tryptophan, permitting further increased delivery into the brain. An expected consequence of increased brain tryptophan levels would be augmented levels of brain quinolinic acid. Moroni et al^{21,22} reported that quinolinic acid levels are increased in the CSF of patients suffering from hepatic encephalopathy or coma and in the brain parenchyma of animals with experimental liver damage.

Bucci et al⁶³ addressed the question of hepatic-related tryptophan neurotoxicity by chronically feeding rats 200 mg tryptophan/kg/d. In normal rats, no changes occurred in CNS morphology, but in rats with portocaval shunts (in which brain tryptophan delivery is further increased, as described above), histologic changes were noted. These changes included an increased number of enlarged astrocytes and neuronal nuclear and process degeneration in a variety of brain regions, including the caudate nucleus.

Other neurologic disorders. Alterations of the metabolism of tryptophan into kynurenines may also occur in a number of other disorders in which neurologic symptoms exist. Included among these are porphyrias, caused by metabolic defects in porphyrin synthesis or breakdown; neurologic sequelae are common. Porphyrin is the cofactor for the 1st enzyme in the kynurenine pathway, tryptophan pyrrolase, and alterations in porphyrin levels or structure could affect the activity and regulation of this enzyme.⁶⁴

In addition, case reports have appeared in the literature that suggest the appearance of transient neurologic and psychiatric symptoms in patients receiving tryptophan doses ranging from 2 to 10 grams. Included among these symptoms are convulsive seizures, anxiety, and hyperirritability.⁶⁵⁻⁶⁷ Other reports indicate behavioral side effects of tryptophan in combination therapy with other pharmaceuticals.⁶⁸⁻⁷⁰ Some studies also suggest an association between inherited disorders of tryptophan metabolism and ataxia and developmental delay.⁷¹⁻⁷⁶

Conclusion. Recent studies have suggested that the synthesis and release of neuroactive substances other

than serotonin may be linked to tryptophan availability. Of these, quinolinic acid is particularly relevant, given its role as a potential neurotoxic agent in a variety of neurologic diseases as well as its agonist properties with respect to the NMDA (AA₁) receptor, implicated in the function of memory and learning through long-term potentiation.⁷⁷ The presence of quinolinic acid in the brain extracellular fluid compartment indicates that, although it is an intermediate metabolite in the intracellular conversion of tryptophan to NAD, quinolinic acid may play additional roles within the nervous system. Thus, it is possible that some behavioral and biochemical effects resulting from experimental alterations in tryptophan levels or metabolism in test animals may be due to changes in kynurenine pathway intermediates, rather than serotonin and its metabolites.

Of great interest is the observation that 2 kynurenine metabolites, quinolinic acid and kynurenic acid, have opposite effects on the NMDA receptor, the former acting as an agonist and the latter as an antagonist.²⁷ Kynurenic acid, therefore, can achieve a neuroprotective effect in the brain in the presence of excitotoxins such as quinolinic acid. However, when compared with quinolinic acid, kynurenic acid levels are far less responsive to acute tryptophan precursor loading; rat brain extracellular levels of kynurenic acid increased less than 10-fold following an intraperitoneal injection of 100 mg/kg tryptophan (unpublished observations). Also of interest, the tryptophan metabolite, serotonin, appears to modulate the sensitivity of the NMDA receptor for its agonists.⁷⁸

Until a better understanding of the interaction among tryptophan, kynurenine pathway intermediates, and the NMDA receptor is achieved, perhaps caution should be exercised with respect to tryptophan self-medication by subsets of the general population.⁷⁹

References

1. Fernstrom JD, Wurtman RJ. Brain serotonin content: physiological dependence on plasma tryptophan levels. *Science* 1971;173:149-152.
2. Moir ATB, Eccleston D. The effects of precursor loading in the cerebral metabolism of 5-hydroxyindoles. *J Neurochem* 1968;15:1093-1108.
3. Hartmann E, Spinweber CL. Sleep induced by L-tryptophan—effect of dosages within the normal dietary intake. *J Nerv Ment Dis* 1979;167:497-499.
4. Hartmann E. Effects of L-tryptophan on sleepiness and on sleep. *J Psychiatric Res* 1983;17:107-113.
5. Hrboticky N, Anderson GH. Effects of L-tryptophan on short term food intake and lean men. *Nutr Res* 1985;5:595-607.
6. Moldofsky H, Luc FA. The relationship of alpha and delta EEG frequencies to pain and mood in "fibrositis" patients with chlorpromazine and L-tryptophan. *Electroencephalogr Clin Neurophysiol* 1980;50:9:71-80.
7. Moller SE, Kirk C, Honore P. Relationship between plasma ratio of tryptophan to competing amino acids and the response to L-tryptophan treatment in endogenously depressed patients. *J Affective Disord* 1980;2:47-59.
8. Sved AF, van Itallie CM, Fernstrom JD. Studies on the antihypertensive action of L-tryptophan. *J Pharmacol Exp Ther* 1982;221:329-332.
9. Wilcock GF, Stevens J, Perkins A. Trazodone/tryptophan for aggressive behaviour. *Lancet* 1987;1:929-930.

10. Salter M, Knowles RG, Pogson CI. Quantification of the importance of individual steps in the control of aromatic amino acid metabolism. *Biochem J* 1986;234:634-647.
11. Gal EM, Sherman AD. L-Kynurenine. Its synthesis and possible regulatory function in brain. *Neurochem Res* 1980;5:223-239.
12. Heyes MP. Hypothesis: a role of quinolinic acid in the neuropathology of glutaric aciduria, type 1. *J Neurol Sci* 1987;14:441-443.
13. Moroni F, Lombardi G, Carla V, Moneti G. The excitotoxin quinolinic acid is present and unevenly distributed in the rat brain. *Brain Res* 1984;295:352-355.
14. Moroni F, Lombardi G, Moneti G, Aldinio C. The excitotoxin quinolinic acid is present in the brain of several mammals and its cortical content increases during the aging process. *Neurosci Lett* 1984;47:51-55.
15. During MJ, Heyes MP, Freese A, et al. Quinolinic acid concentrations in striatal extracellular fluid reach potentially neurotoxic levels following systemic L-tryptophan loading. *Brain Res* 1988;476:384-387.
16. During MJ, Freese A, Heyes MP, et al. Neuroactive metabolites of L-tryptophan, serotonin and quinolinic acid, in striatal extracellular fluid: effect of tryptophan loading. *FEBS Lett* 1989;247:438-444.
17. Stone TW, Connick JH. Quinolinic acid and other kynurenines in the central nervous system. *Neuroscience* 1985;15:597-617.
18. Ben-Ari Y. Limbic seizures and brain damage produced by kainic acid: mechanisms and relevance to human temporal lobe epilepsy. *Neuroscience* 1985;14:375-403.
19. Zaczek R, Coyle JT. Excitatory amino acid analogues: neurotoxicity and seizures. *Neuropharmacology* 1982;21:15-26.
20. Lapin IP. Kynurenines and seizures. *Epilepsia* 1981;22:257-265.
21. Moroni F, Lombardi G, Carla V, Cal S, Etienne P, Nair NPV. Increase in the content of quinolinic acid in cerebrospinal fluid and frontal cortex of patients with hepatic failure. *J Neurochem* 1986;47:1667-1671.
22. Moroni F, Lombardi G, Carla V, Pellegrini D, Carassale GL, Cortesini C. Content of quinolinic acid and other tryptophan metabolites increases in brain regions of rats used as experimental models of hepatic encephalopathy. *J Neurochem* 1986;46:869-874.
23. Beal MF, Kowall NW, Ellison DW, Mazurek MF, Swartz KJ, Martin JB. Replication of the neurochemical characteristics of Huntington's disease by quinolinic acid. *Nature* 1986;321:168-171.
24. Whetsell WO Jr. The use of organotypic tissue culture for the study of amino acid neurotoxicity and its antagonists in the mammalian CNS. *Clin Neuropharmacol* 1984;7:248-250.
25. McLennan H. Receptors for excitatory amino acids in the mammalian central nervous system. *Prog Neurobiol* 1983;20:251-271.
26. Perkins MN, Stone TW. Pharmacology and regional variations of quinolinic acid-evoked excitations in the rat central nervous system. *J Pharmacol Exp Ther* 1983;226:551-557.
27. Stone TW, Burton NR. NMDA receptors and ligands in the vertebrate CNS. *Prog Neurobiol* 1988;30:333-368.
28. Foster AC, Fagg GE. Acidic amino acid binding sites in mammalian neuronal membranes: their characteristics and relationship to synaptic receptors. *Brain Res Rev* 1984;7:103-164.
29. Perkins MN, Stone TW. Quinolinic acid: regional variations in neuronal sensitivity. *Brain Res* 1983;259:172-176.
30. Schwarcz R, Whetsell WO, Mangano RM. Quinolinic acid: an endogenous metabolite that produces axon-sparing lesions in the rat striatum. *Science* 1983;219:316-318.
31. Choi DW. Dextrorphan and dextromethorphan attenuate glutamate neurotoxicity. *Brain Res* 1987;403:333-336.
32. Foster AC, Gill R, Kemp JA, Woodruff GN. Systemic administration of MK-801 prevents N-methyl-D-aspartate induced neuronal degeneration in rat brain. *Neurosci Lett* 1987;76:307-311.
33. Beal MF, Kowall NW, Swartz KJ, Ferrante RJ, Martin JB. Systemic approaches to modifying quinolinic acid lesions in rats. *J Neurosci* 1988;8:3901-3908.
34. Schwarcz R, Meldrum B. Excitatory amino acid antagonists provide a novel therapeutic approach to neurological disorders. *Lancet* 1985;2:140-143.
35. Gholson RK, Ueda I, Ogasawara N, Henderson LM. The enzymatic conversion of quinolinic acid to nicotinic acid mononucleotide in mammalian liver. *J Biol Chem* 1964;239:1208-1214.
36. Foster AC, Zinkand WC, Schwarcz R. Quinolinic acid phosphoribosyltransferase in rat brain. *J Neurochem* 1985;44:446-454.
37. Foster AC, Schwarcz R. Characterization of quinolinic acid phosphoribosyltransferase in human blood and observations in Huntington's disease. *J Neurochem* 1985;45:199-205.
38. Perez-Cruet J, Chase TN, Murphy DL. Dietary regulation of brain tryptophan metabolism by plasma ratio of free tryptophan and neutral amino acids in humans. *Nature* 1974;248:693-695.
39. McMenamy RH, Oncley JL. The specific binding of L-tryptophan to serum albumin. *J Biol Chem* 1958;233:1436-1447.
40. Knott PJ, Curzon G. Free tryptophan in plasma and brain tryptophan metabolism. *Nature* 1972;239:452-453.
41. Pardridge WM. Blood-brain transport of nutrients: introduction. *Fed Proc* 1986;45:2047-2049.
42. Fernstrom JD, Wurtman RJ. Brain serotonin content: increase following ingestion of carbohydrate diet. *Science* 1971;174:1023-1025.
43. Belendiuk K, Belendiuk GW, Freedman DX. Blood monoamine metabolism in Huntington's disease. *Arch Gen Psychiatry* 1980;37:325-332.
44. Watt JAG, Cunningham WL. Plasma amino acid levels in Huntington's chorea. *Br J Psychiatry* 1978;132:394-397.
45. Perry TL, Diamond S, Hansen S, Stedman D. Plasma amino acid levels in Huntington's chorea. *Lancet* 1969;1:806-808.
46. Barbeau A. L-Dopa and juvenile Huntington's disease. *Lancet* 1979;2:1066.
47. McLeod WR, Horne DJ de L. Huntington's chorea and tryptophan. *J Neurol Neurosurg Psychiatry* 1972;32:510-513.
48. Oepen I, Oepen H. Tryptophanbelastungstest bei Huntingtontschorea. *Humangenetik* 1969;7:197-202.
49. Oliphant J, Evans JI, Forrest AD. Huntington's chorea—some biochemical and therapeutic aspects. *J Ment Sci* 1960;106:718-725.
50. Schwarcz R, Okuno E, White RJ, Bird ED, Whetsell WO. 3-Hydroxyanthranilate oxygenase activity is increased in the brains of Huntington disease victims. *Proc Natl Acad Sci USA* 1988;85:4079-4081.
51. Foster AC, Whetsell WO, Bird ED, Schwarcz R. Quinolinic acid phosphoribosyltransferase in human and rat brain: activity in Huntington's disease and in quinolinic acid lesioned rat striatum. *Brain Res* 1985;336:207-214.
52. Aronin N, Cooper PE, Lorenz LJ, et al. Somatostatin is increased in the basal ganglia in Huntington disease. *Ann Neurol* 1983;13:519-526.
53. Ferrante RJ, Kowall NW, Beal MF, Richardson EP, Martin JB. Selective sparing of a class of neurons in Huntington's disease. *Science* 1985;230:561-563.
54. Mans AM, Saunders SJ, Kirsch RE, Biebuyck JF. Correlation of plasma and brain amino acid and putative neurotransmitter alterations during acute hepatic coma in the rat. *J Neurochem* 1979;32:285-292.
55. Young SN, Lal S, Sourkes TL, Feldmuller F, Aronoff A, Martin JB. Relationship between tryptophan in serum and CSF, and 5-hydroxyindoleacetic acid in CSF of man: effect of cirrhosis of liver and probenecid administration. *J Neurol Neurosurg Psychiatry* 1975;38:322-330.
56. Hirayama C. Tryptophan metabolism in liver disease. *Clin Chim Acta* 1971;32:191-197.
57. Curzon G, Kantamenini BD, Fernando JC, Woods MS, Cavanagh JB. Effects of chronic portocaval anastomosis on brain tryptophan, tyrosine and 5-hydroxytryptophan. *J Neurochem* 1975;24:1065-1070.
58. Smith A, Rossi-Fanelli F, Ziparo V, James JH, Perelle BA, Fischer JE. Alterations in plasma and CSF amino acids, amines, and metabolites in hepatic coma. *Ann Surg* 1978;187:343-350.
59. Munro HN, Fernstrom JD, Wurtman RJ. Plasma neutral amino acids and tryptophan in cirrhosis. *Lancet* 1975;2:419.
60. Soeters PB, Fischer JE. Insulin, glucagon, amino acid imbalance, and hepatic encephalopathy. *Lancet* 1976;2:880-882.
61. Grippon P, Lafitte MLP, Boschall M, et al. Evidence for the role of ammonia in the intracerebral transfer and metabolism of tryptophan. *Hepatology* 1986;6:682-686.
62. Curzon G, Knott PJ. Environmental toxicological, and related aspects of tryptophan metabolism with particular reference to the central nervous system. *CRC Crit Rev Toxicol* 1977;5:145-187.
63. Bucci L, Ioppolo A, Chiavarelli R., Biogotti A. The central nervous system toxicity of long-term oral administration of L-tryptophan

- to porto-caval shunted rats. *Br J Exp Pathol* 1982;63:235-241.
64. Litman DA, Lorreia MA. L-Tryptophan: a common denominator of biochemical and neurological events of acute hepatic porphyria? *Science* 1983;222:1031-1033.
 65. Coursin DB. Recommendations for standardization of the tryptophan load test. *Am J Clin Nutr* 1964;14:56-61.
 66. Coursin DB. Vitamin B6 metabolism in infants and children. *Vitam Horm* 1964;22:755-786.
 67. Coursin B. Central nervous system hypersensitivity to tryptophan. *Am J Clin Nutr* 1971;24:821-825.
 68. Steiner W, Fontaine R. Toxic reaction following the combined administration of fluoxetine and L-tryptophan: five case reports. *Biol Psychiatry* 1986;21:1067-1071.
 69. Levy AB, Bucher P, Vtolato N. Myoclonus, hyperreflexia, and diaphoresis in patients on phenelzine-tryptophan combination treatment. *Can J Psychiatry* 1985;30:434-436.
 70. Hall CD, Weiss EA, Morris CE, Prange AJ. Rapid deterioration in patients with parkinsonism following tryptophan-pyridoxine administration. *Neurology* 1972;22:231-237.
 71. Tada K, Ito H, Wada Y, Arakawa T. Congenital tryptophanuria with dwarfism. *Tohoku J Exp Med* 1963;80:118-121.
 72. Wong PWK, Forman P, Tabahoff B, Justice P. A defect in tryptophan metabolism. *Pediatr Res* 1976;10:725-729.
 73. Komrower GM, Wilson V, Clamp JR, Westall RG. Hydroxykynureninuria. A case of abnormal tryptophan metabolism probably due to a deficiency of kynureninase. *Arch Dis Child* 1964;39:250-254.
 74. Price JM, Yess N, Brown RR, Johnson SAM. Tryptophan metabolism. A hitherto unreported abnormality occurring in a family. *Arch Derm* 1967;95:462-467.
 75. Salih MAM, Bender DA, McCreanor GM. Lethal familial pellagra-like skin lesion associated with neurological and developmental impairment and the development of cataracts. *Pediatrics* 1985;76:787-791.
 76. Fenton DA, Wilkinson JD, Toseland PA. Family exhibiting cerebellar-like ataxia, photosensitivity and shortness of stature—a new inborn error of tryptophan metabolism. *J Roy Soc Med* 1983;76:736-740.
 77. Danysz W, Wroblewski JT, Costa E. Learning impairment in rats by N-methyl-D-aspartate receptor antagonists. *Neuropharmacology* 1988;27:653-656.
 78. Reynolds JN, Baskys A, Carlen PL. Serotonin enhances responses to N-methyl-D-aspartate in rat neocortical neurons. 17th Annual Meeting Soc Neurosci 1987;Abstract 433.11.
 79. Freese A, Swartz KJ, During MJ. Potential neurotoxicity of tryptophan. *Ann Intern Med* 1988;108:312-313.